Comparative analysis of incidence of leptospirosis among farm animals and humans in Ukraine


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Abstract

Leptospirosis remains one of the most widespread natural-focal, zoonotic infectious diseases in the world and in Ukraine. Leptospirosis is enzootic in the entire territory of Ukraine. Cases of diseases are registered in all regions of Ukraine. We initiated a study of comparative analysis of territorial distribution of leptospirosis outbreaks among animals and incidence in humans in Ukraine covering the years 2009–2016 inclusive. This study of the incidence of leptospirosis in Ukraine shows a significant circulation of leptospirosis both among humans and animals. Among cattle herds in Ukraine the percentage of positive animals was found to be 4.2% of the surveyed population. The dominant serovars of Leptospira were khabara (12.4%) and polonica (9.5%). Positive reactions with other serovars were observed less frequently: tarassovi – 5.1%, bratislava – 4.9%, copenhageni – 4.1%, grippotyphosa – 2.4%, pomona – 1.1%, canicola – 1.0%. In pigs, the percentage of positive animals amounted to 3.2%, the dominant serovars of Leptospira were bratislava (29.1%) and copenhageni (25.1%). Positive reactions with other serovars were observed less frequently: tarassovi – 4.3%, canicola – 3.0%, pomona – 2.7%, grippotyphosa – 1.3%, polonica – 1.2%, khabara – 0.6%. In horses, the percentage of positive animals amounted to 9.5% of the surveyed population. The serological range of Leptospira in horses was as follows: copenhageni – 14.2%, bratislava – 12.1%, canicola – 6.8%, grippotyphosa – 4.8%, tarassovi – 4.7%, pomona – 2.1%, khabara – 1.4%, polonica – 1.3%. Analysis of the results of research indicates extensive circulation of leptospirosis among humans in Ukraine as evidenced by the percentage of humans positively responding to MAT – 12.1% of the studied samples. The etiological structure of leptospirosis cases includes all the 14 serovars of the diagnostic set. The basis of the etiological spectrum was the serovar copenhageni – 37.3%. The share of other serovars as the etiological factor of leptospirosis in humans was different in different spans of the considered period. Most frequently, those were khabara – 12.3%, grippotyphosa – 11.7%, canicola – 9.5%, pomona – 9.1%. We mapped annual incidence of leptospirosis in animals and humans. Choropleth maps of annual leptospirosis incidence and cluster maps show opposite spatial patterns for animals and humans. The highest human rates were in the western and central parts of the country while the highest animal rates were mainly in the eastern part.

Keywords: Leptospira; etiological structure; microscopic agglutination test; mapping; GIS.

Introduction

Leptospirosis is an infectious disease that affects a large number of mammal species, as well as humans (Ko et al., 2009; Sykes et al., 2011). It has become widespread in many countries around the world (Levett, 2001; Lee et al., 2017; Daud et al., 2018). Leptospirosis is one of the most common and significant natural foci zoonoses and has been diagnosed on five continents (except Antarctica) in most countries of the world (Bharti et al., 2003). Leptospirosis causes huge economic losses in animal husbandry because of its significant incidence among animals, leading to mass abortions and a large number of still births, a decrease in animal productivity, and significant costs incurred in diagnostic research, treatment and prophylactic and quarantine measures (Hartskeerl et al., 2011; Pavlenko et al., 2011). Leptospirosis is a disease of great social significance because leptospirosis infected animals and host animals pose a direct threat to public health, characterized by high mortality of patients in recent years (Santos et al., 2018). Leptospirosis has been reported in over 150 mammalian species (Malalana et al., 2017; Brown et al., 2018; Miyama et al., 2018). In addition, antibodies against Leptospira strains can be identified in the blood sera of reptiles and amphibians (Levett, 2001; Adler & Moctezuma, 2010). Among farm animals, this zoonosis is often registered in cattle (Alonso-Andicoberry et al., 2001; Talpada et al., 2006), swine (Hartleben et al., 2013; Pyskun et al., 2016; Bertollini et al., 2018) and horses (Brem et al., 1999; Arent & Kędzierska-Mieszkowska, 2013; Arent et al., 2016) throughout the world.

The majority of the leading specialists in the study of leptospirosis have studied the etiological structure of the disease, which of course, requires the constant monitoring both farm animals and dogs, synanthropic rodents and residents of natural foci (Stepna et al., 2016). At this stage, one of the main tasks of epidemiological and epizootic monitoring of natural focal infections is to determine enzootic areas of circulation of pathogens, including leptospirosis (Pavlenko et al., 2011).

Analyses of pathogenic Leptospira isolated in different countries shows that the etiological structure of leptospirosis in specific areas (dis-
tricts, regions, and countries) is heterogeneous in the number of various serotypes of *Leptospira*, and their ratios. The objective of our study was comparative analysis of the territorial confines of leptospirosis outbreaks among animals and its incidence in humans in Ukraine.

**Materials and methods**

Analysis of the etiological structure of leptospirosis based on data from animals. The spread and etiological structure of leptospirosis in animals was analyzed according to reports of the State Scientific Research Institute of Laboratory Diagnostics and Veterinary Sanitary Expertise during the years 2009–2016. Sero-prevalence for each region was calculated as the number of leptospirosis positive samples divided by the sample quantity in the region. Calculation of exact binomial 95% confidence intervals (BCI) was performed for sero-prevalence estimates using the R package (https://cran.r-project.org).

Antigens. Cultures of reference strains of *Leptospira* consisting of 8 serovars: canicola, grippotyphosa, kubura, copenhageni, pomona, polonica, tarassovi and bratislava, were used to perform the microscopic agglutination test (MAT). These diagnostic strains of *Leptospira* used in serological studies on leptospirosis of animals were prepared by veterinary diagnostic laboratories in Ukraine. Leptospira were cultivated in Korthof liquid medium at 28–30 °C under aerobic conditions. The strains were subcultured every 7–10 days.

Microscopic agglutination test. The test was carried out according to OIE Manual of Standards for Diagnostic Tests and Vaccines and under the current Regulations (Nastanova z laboratornoi diagnostichnoi leptospirozi [Guidance on laboratory diagnosis of leptospirosis]. (1997). Zaraiestrovana 11.01.1997, No 15–14/2. Ministerstvo Silskoho Hospodarstva i Prodovoltva Ukrainy, Kyiv). Briefly, the serum samples diluted 1 : 25 were mixed with an equal volume of each of the *Leptospira* culture. Serum dilution (including added antigen) during preliminary examination was 1 : 50. For samples reacting in the preliminary examination with one or more serovars, series of twofold dilutions were prepared to reach the end point – 50% agglutination. The samples with titers equal or higher than 1 : 50 were recognized as positive.

Analysis of etiological structure of leptospirosis based on human data. Spreading and etiological structure of leptospirosis in humans and annual human incidence data per 100,000 population was analyzed according to reports of the State Institution Ukrainian Center for Diseases Control and Monitoring of the Ministry of Health of Ukraine during the years 2009–2016.

Antigens. Cultures of reference strains of *Leptospira* consisted of 14 serovars: canicola, grippotyphosa, kubura, copenhageni, pomona, polonica, tarassovi, bratislava, javonica, autumnalis, djatti, balantium, pyrogens and cynopteri, were used to perform the MAT. These diagnostic strains of leptospires are used in serological studies on leptospirosis of animals that are conducted by the State Institution Regional Laboratory Center Ministry of Health of Ukraine.

Microscopic agglutination test. The test was carried out under the current Regulations (Protevpidemichni zakhody ta laboratorna diagnostyka leptospirozi [Preventive measures and laboratory diagnostics of leptospirosis. Methodological instructions. MI 9.1.109–02]. (2002). Zatverzhdena 11.12.2002, No 39, Kyiv). The serum samples diluted 1 : 50 were mixed with an equal volume of each of the *Leptospira* serovars. Final serum dilution (including added antigen) during preliminary examination was 1 : 100. For samples reacting in the preliminary examination with one or more serovars, a series of twofold dilutions were prepared to reach the end point – 50% of agglutination. The samples with titers equal or higher than 1 : 100 were recognized as positive.

To calculate annual incidence per 100,000 animals, population data from the web site (www.ukrstat.gov.ua) of the State Statistics Service of Ukraine were used. The data on all three farm animal species (cattle, swine, horses) were summarized.

Annual incidence by region of both animals and humans was choropleth mapped in ESRI ArcGIS 10.3. Quantile classification with 5 classes of the data was chosen. With this classification, an equal number of regions fall into each class.

To find out and compare the patterns of incidence distribution among animals and humans at the regional level, we applied the local Moran I statistics at a statistical significance P < 0.05 using 999 permutations in the software GeoDa 1.0.0.8. In the analysis, we applied the first order rook contiguity matrix, where the regions adjoining their boundaries are considered neighbours. Local Moran I statistics reveals both clusters of similar values, and areas dissimilar to their neighbours – spatial outliers. On the maps B, areas with a high incidence rate, surrounded by high-value areas are designated High-High, areas with a low value surrounded by areas with a low values – Low-Low. Areas with a high incidence of disease surrounded by neighbours with low values are designated High-Low, areas with a low incidence rate surrounded by high values are Low-High.

**Results**

During the years 2009–2016, 1,238,876 samples of cattle sera were investigated by veterinary diagnostic laboratories of Ukraine and 52,310 reacted positive for leptospirosis. Analysis of the results indicates extensive circulation of leptospirosis among cattle herds in Ukraine as evidenced by the percentage of cattle positively responding to MAT, which is 4.2% (BCI, 4.2–4.3%) of the studied samples.

![Fig. 1. Etiological structure of leptospirosis in cattle in Ukraine (2009–2016 years)](image)

As shown in Figure 1 the dominant serovars of *Leptospira* were kubara (12.4%; BCI, 12.1–12.7%), and polonica (9.5%; BCI, 9.2–9.7%). Positive reactions with other serovars were observed less frequently: tarassovi – 5.1% (BCI, 4.9–5.3%), bratislava – 4.9% (BCI, 4.7–5.1%), copenhageni – 4.1% (BCI, 4.0–4.3%), grippotyphosa – 2.4% (BCI, 2.3–2.6%), pomona – 1.1% (BCI, 1.0–1.2%), copenhageni – 1.0% (BCI, 0.9–1.0%). The proportion of the cattle disease cases where antibodies to multiple serovars of *Leptospira* (cross tests) were detected was 59.6% (BCI, 58.9–60.2%).

During this period, 989,659 samples of pigs’ sera were tested by veterinary laboratories in Ukraine and 31,181 samples were regarded as positive for leptospirosis, which amounted to 3.2% (BCI, 3.1–3.3%) of the surveyed population.

![Fig. 2. Etiological structure of leptospirosis in pigs in Ukraine (2009–2016 years)](image)

As shown in Figure 2, the dominant serovars of *Leptospira* were bratislava (29.1%; BCI, 28.5–29.7%) and copenhageni (25.1%; BCI,
were examined in Ukraine and 6,734 samples were regarded as positive. The proportion of the pigs' disease cases where antibodies to multiple serovars of Leptospira were detected was 32.8% (BCI, 32.1–33.4%).

During the years 2009–2016, 76,674 samples of sera from horses were examined in Ukraine and 6,734 samples were regarded as positive for leptospirosis. Analysis of the results of testing, indicates extensive circulation of leptospirosis among horses in Ukraine, which amounted to 9.5% (BCI, 9.3–9.8%) of the surveyed population. Most frequently, those were kobura – 12.3% (BCI, 11.0–13.6%), grippotyphosa – 11.7% (BCI, 10.5–13.0%), canicola – 9.5% (BCI, 8.5–10.7%), pomona – 9.1% (BCI, 8.0–10.2%). Positive reactions with other serovars were observed less frequently: bratislava – 4.8% (BCI, 3.7–5.2%), javanica – 4.0% (BCI, 3.3–4.8%), autumnalis – 2.7% (BCI, 2.1–3.3%), copenhageni – 2.2% (BCI, 1.7–2.8%), ballum – 2.1% (BCI, 1.6–2.6%), tarassovi – 2.0% (BCI, 1.6–2.6%), djatzi – 1.4% (BCI, 1.0–1.9%), pyrogenes – 0.8% (BCI, 0.5–1.2%), polonica – 0.6% (BCI, 0.3–0.9%) (Fig. 5).

Discussion

This study shows a significant circulation of leptospirosis both among humans and animals in Ukraine. Among cattle herds, the percentage of reacting positive animals is 4.2% (BCI, 4.2–4.3%), in pigs it amounted to 3.2% (BCI, 3.1–3.2%) and in horses it amounted to 9.5% (BCI, 9.3–9.8%) of the surveyed population. Analysis of the results of research indicates extensive circulation of leptospirosis among humans in Ukraine as evidenced by the percentage of positively responding to MAT humans – 12.1% (BCI, 11.7–12.5%) of the studied samples. It was established that the epizootic situation concerning animal leptospirosis and infection in animals in Ukraine (2009–2016 years)
incidence of leptospirosis in humans in different regions of Ukraine differ both in the etiological structure of agents, and the number of cases. During the analysis of the leptospirosis circulation, we examined for the period 2009–2016 a large number of specimens from animals and people from all over the country: cattle – 1,238,876 samples, pigs – 989,659, horses – 70,674 samples of sera and 24,990 samples of sera of humans.

Fig. 7. Distribution of animal and human leptospirosis cases in Ukraine (2009 year): a – animal and human leptospirosis incidence per 100,000 at region level, b – cluster and outlier maps of animal and human leptospirosis incidence

Fig. 8. Distribution of animal and human leptospirosis cases in Ukraine (2010 year): a – animal and human leptospirosis incidence per 100,000 at region level, b – cluster and outlier maps of animal and human leptospirosis incidence
Regarding the limitations of our research, it should be noted that we did not take into account the distribution and etiological structure of leptospirosis in dogs in Ukraine, since studies of leptospirosis in dogs are conducted mainly on the request of private veterinary clinics, and there is no official state programme for monitoring this disease in dogs. At the same time, leptospirosis in dogs constitutes a significant threat of transmission of leptospirosis to humans (Whitney et al., 2009; Sykes et al., 2011). Leptospirosis is a zoonosis, so cases of this disease in people are observed in areas where there are animals sick with leptospirosis or there are animal hosts (Witkowski et al., 2016; Guernier et al., 2017). Choropleth maps of annual leptospirosis incidence and cluster maps show opposite spatial patterns for animals and humans (Fig. 7–14). The highest human rates are in western and central parts of the country while the highest animal rates are mainly in the eastern part.
In our opinion, there are two main reasons for the opposite spatial patterns for animals and humans. The first reason is the difference in the causes of the spread of leptospirosis in humans and animals (Flores et al., 2017; Rajala et al., 2017). The most important reasons for the widespread distribution of leptospirosis in animals in Ukraine are inadequate animal welfare, and frequent and uncontrolled movement of them from farm to farm (Malakhov et al., 2000; Sykes et al., 2011). At the same time, natural conditions and rodent population have a secondary role in the occurrence of leptospirosis among animals. In contrast to animals, the main cause of infection in humans are rodents (rats and mice) (Stepna et al., 2016; Santos et al., 2018), insufficient rodent control and natural conditions. The main path of infection with *Leptospira* in Ukraine to humans, in contrast to leptospirosis of animals, is through water (swimming, fishing, working in wet areas, etc.) (Whitney et al., 2009; Barragan et al., 2016; Pinto et al., 2017). The proportion of people with leptospirosis associated with reservoirs and wet places is 50.7%; transmission through food related to the objects of epidemic – 2.2%; contact and household, connected with settlements – 31.9%; not established – 15.2% (Information report on the epidemic situation with leptospirosis in Ukraine in 2016).
Fig. 13. Distribution of animal and human leptospirosis cases in Ukraine (2015 year): a – animal and human leptospirosis incidence per 100,000 at region level, b – cluster and outlier maps of animal and human leptospirosis incidence

Fig. 14. Distribution of animal and human leptospirosis cases in Ukraine (2016 year): a – animal and human leptospirosis incidence per 100,000 at region level, b – cluster and outlier maps of animal and human leptospirosis incidence

The second reason, in our opinion, is underestimation and possible underreporting of cases in animals and insufficient monitoring of leptospirosis in animals. This applies particularly to the western regions of Ukraine (Ukhovskiy et al., 2015).

Conclusion

Based on the results of monitoring studies of the territory of Ukraine, we found that leptospirosis is widespread both among livestock and also among people. The average sero-prevalence for 8 years (2009–2016) was: among cattle herds – 4.2% of the surveyed population, in pigs up to 3.2%, in horses up to 9.5% and in humans – 12.1% of the studied samples. It was determined that the spectrum of leptospirosis of agricultural animals and humans in Ukraine had peculiarities both within the range of serovars and their significance for the total pathology of the disease. The dominant serovar *Leptospira* circulating within the territory of Ukraine were: in cattle – *kabura* and *polonica*, in pigs – *copenhagenii* and *bratislava*, in horses – *copenhagenii*, *bratislava* and *canicola*, in
humans — *copenhagenii*. Based on the results of a comparative analysis of incidence of leptospirosis among farm animals and humans in the territory of Ukraine, it was established that animals and humans manifest opposite patterns. The highest animal leptospirosis rates were detected mainly in the eastern part while the highest human leptospirosis rates were in the western and central parts of the country. This information on the distribution of leptospirosis in animals and humans can help to improve and optimize the planning and development of specific preventive measures against leptospirosis in Ukraine.

References


