Hereditary tubulopathies including the associated bone disease

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Introduction

Tubulopathies form a heterogeneous group of diseases combined by the presence of disorders in the tubular epithelium of the nephron functions of one or more enzyme proteins that cease to function as reabsorption of one or several substances filtered from the blood through the glomeruli into tubules, which determines the development of the disease. By origin they are classified into primary and secondary tubulopathies. The primary ones involve a hereditary defect of the genes that regulate the function of a particular tubular enzyme, resulting in the development of the disease. This review addresses the tubulopathies accompanying bone disease, namely: de Tony-Debre-Fanconi syndrome (autosomal dominant, autosomal recessive, X-linked), renal distal tubular metabolic acidosis type I (classic, autosomal dominant, autosomal recessive inheritance), renal distal tubular metabolic acidosis I (autosomal dominant, autosomal recessive inheritance) and type II (autosomal recessive inheritance accompanying delayed mental development and eye disorders), combined distal and proximal renal tubular metabolic acidosis type III (autosomal recessive inheritance characterized by osteoporosis), hypophosphatemia rickets (X-linked dominant, autosomal dominant, primary hypercalciuria, autosomal recessive inheritance). However, the diagnosis of tubulopathy remains complex and requires expensive laboratory equipment and specialist expertise; it can be diagnosed in children showing the following symptoms: impaired growth, vitamin D resistant rickets (lower limb deformities between 2 and 3 years of age). In the evaluation of such patients urine analysis is commonly used (levels of calcium, phosphorus, pH, bicarbonate, sodium, potassium, glucose, creatinine, protein, amino acids), blood count (levels of creatinine, uric acid, alkaline phosphatase, glucose, pH and sodium, bicarbonate, potassium, chloride, calcium, phosphorus ions), ultrasound of the kidneys to detect nephrocalcinosis. Determination of serum parathyroid hormone concentration, vitamin D metabolites, aldosterone and plasma renin activity, cysteine lympoocyte concentration (suspicion to diagnose cystinosis) and ophthalmologist examination may also be used as additional diagnostic methods. Despite the fact that most tubulopathies can be diagnosed clinically, molecular genetic studies are needed to clarify the type of inheritance and prognosis. The use of calcitriol will help in the management of phosphorous levels in the blood. Correction of vitamin D deficiency state is not required. Calcitriol supplementation may prevent secondary hyperparathyroidism resulting from increased phosphate intake.

Keywords: vitamin D resistant rickets; children; hypophosphatemia; genes

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Tubulopathy is a heterogeneous group of diseases combined by the nephron functions disorders of one or more enzyme proteins in the tubular epithelium that cease to function as a reabsorption of one or several substances filtered from the blood through the glomeruli into tubules, which determines the development of the disease. This review addresses the tubulopathies accompanying bone disease, namely: de Tony-Debre-Fanconi syndrome (autosomal dominant, autosomal recessive, X-linked), renal distal tubular metabolic acidosis type I (classic, autosomal dominant, autosomal recessive inheritance), renal distal tubular metabolic acidosis I (autosomal dominant, autosomal recessive inheritance) and type II (autosomal recessive inheritance accompanying delayed mental development and eye disorders), combined distal and proximal renal tubular metabolic acidosis type III (autosomal recessive inheritance characterized by osteoporosis), hypophosphatemia rickets (X-linked dominant, autosomal dominant, primary hypercalciuria, autosomal recessive inheritance). However, the diagnosis of tubulopathy remains complex and requires expensive laboratory equipment and specialist expertise; it can be diagnosed in children showing the following symptoms: impaired growth, vitamin D resistant rickets (lower limb deformities between 2 and 3 years of age). In the evaluation of such patients urine analysis is commonly used (levels of calcium, phosphorus, pH, bicarbonate, sodium, potassium, glucose, creatinine, protein, amino acids), blood count (levels of creatinine, uric acid, alkaline phosphatase, glucose, pH and sodium, bicarbonate, potassium, chloride, calcium, phosphorus ions), ultrasound of the kidneys to detect nephrocalcinosis. Determination of serum parathyroid hormone concentration, vitamin D metabolites, aldosterone and plasma renin activity, cysteine lympoocyte concentration (suspicion to diagnose cystinosis) and ophthalmologist examination may also be used as additional diagnostic methods. Despite the fact that most tubulopathies can be diagnosed clinically, molecular genetic studies are needed to clarify the type of inheritance and prognosis. The use of calcitriol will help in the management of phosphorous levels in the blood. Correction of vitamin D deficiency state is not required. Calcitriol supplementation may prevent secondary hyperparathyroidism resulting from increased phosphate intake.

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We provide a brief overview of the tubulopathies associated with bone disease.

**Primary de Toni-Debre-Fanconi syndrome (OMIM 134600, 613388, 615605, 616026)**

Syn.: Fanconi syndrome, glucosaminophosphate diabetes, glucose-phosphate diabetest, rickets hereditary vitamin D-resistant, idiopathic Fanconi syndrome, Fanconi hereditary syndrome, Renal Fanconi syndrome, Fanconi syndrome, Primary de Toni-Debre-Fanconi syndrome, Inherited Fanconi syndrome.

Fanconi syndrome is divided into four types: type I (OMIM 134600), type II (OMIM 613388), type III (OMIM 615605), type IV diabetes with MODY (OMIM 616026).

In its complete form, Fanconi syndrome (disease) is characterized by a triad of symptoms: hypophosphatemia associated with bone disease, excessive renal wasting of glucose and amino acids, arising as a result of violations in the proximal segment of the nephron (Bacconi et al., 2005). It is associated with the most severe types of prolonged tubulopathies. The disease is genetically predisposed (Bai et al., 2004; White et al., 2005).

Among other researchers, a Swiss pediatrician Fanconi was the first to describe the particular signs of the disease. In 1931, he described a child with dwarfism, rickets, glycosuria and albuminuria. Two years later, de Toni found hypophosphatemia to be further clinical evidence and, later on, Debré determined elevated levels of organic acids in urine, called aminoaciduria (Younes et al., 2003; Zvičnja et al., 2011).

**Aetiopathogenesis**

It is believed that the genetically determined defects of enzymatic phosphorylation in the renal tubules (combined tubulopathy); deficiency of enzymes from the complexes II and III (succinate dehydrogenase and cytochrome oxidase) of the respiratory chain are the basis of the disease. Some authors believe that the basis of the disease is mitochondrial genesis (Lichter-Konecki et al., 2001; Watanabe, 2017).

These mutations lead to various defects in the renal proximal tubules leading to excessive urinary waste of phosphates, glucose and amino acids, as well as to the acid-base imbalance. Metabolic acidosis and insufficiency of phosphorus compounds can also contribute to bone deformities (ostemalacia (adults) and rickets (children)) (Pesik et al., 2015).

Based on its etiology, the syndrome can be divided into two main categories: primary (hereditary) and secondary (acquired). The secondary one is the most common.

The primary (hereditary) syndrome resulting from a genetic mutation occurs in approximately 1 in 20,000 births, de Tony-Debre-Fanconi disease occurs in approximately 1 in 40,000 births. This disease is believed to be caused by the damage to the transport systems in the proximal tubules resulting in disruption of phosphate, glucose and amino acids transportation. Typical episodic features include dehydration, symptoms of rickets and delayed growth. Sometimes the disorder manifests itself at an older age as a renal failure (Tasic, 2008; Besouw et al., 2017). Damage to the sodium transport systems in the proximal tubules (for example, in acute renal failure) leads to a pronounced sodium reabsorption and tubular acidosis disorder, hydrogen ion transport and proximal tubules reabsorbed substances can be disrupted: glucose, phosphate, uric acid, amino acids.

An acquired Fanconi syndrome develops associated with other hereditary disorders or kidney diseases, namely: congenital metabolism or transport disorders (cystitis, tyrosinemia type I, glucogenosis type XI, galactosaemia, congenital intolerance to fructose, Wilson disease, ociclocerebrorenal syndrome (Lowe syndrome), vitamin D-resistant rickets, impaired energy metabolism, Macardle-Schmid-Pearson disease, cyclic citrate C oxidase deficiency (COX deficiency), pyruvate carboxylase (PC) deficiency, carnitine palmitoyltransferase I (CPT I) deficiency); chronic diseases (paraproteinemias (multiple myeloma), tubulointerstitial nephropathies, nephrotic syndrome, nephropathy in renal transplant allografts, malignant tumors (paraneoplastic disease)); heavy metal salts intoxication (mercury, lead, cadmium, uranium); organophosphate poisoning (toluene, maleic acid, lysol); drug-induced toxicity (platinum-based agents, expired tetracycline and gentamycin), severe burns. The syndrome may be complete if none of these three symptoms (glycosuria, phosphaturia and aminoaciduria) are observed and incomplete if there are only two of them:

- glucosaminophosphate diabetes without acidosis (described by Dent and Kyle);
- phosphoglucone diabetes (described by Mac Cune);
- aminoacidic diabetes (described by Jornvat, Wallgren and Nicola); glucosamine diabetes (described by Julliard and Fischer).

**Clinical evidence**

The severity of clinical manifestations and metabolic disorders may differ depending on two clinical and biochemical variants of the disease: in children (early) and adults (late). The pediatric form arises during the 1st year of life and manifestations can include frequent vomiting, loss of appetite, mental and physical developmental delay, a tendency to severe infectious diseases. Gradually there is a proportional dwarfism, rickets and renal insufficiency.

Early dwarfism: intense increase in the rate of growth in height and weight (up to 30%) that occurs during the 5–6 of normal growth and weight gain.

Median age at diagnosis of rickets is 10–12 months; it is characterized by a topographical specificity: the skull is deformed by localized impact; in contrast to fractures in the thoracic (mid back) spine and limbs. Bone pain of moderate intensity primarily tends to be localized in the limbs and spine. It is associated with severe hypocalcemia (1.6–1.8 mmol/L) and reduced intestinal calcium absorption.

Polydipsia and polyuria are common for the beginning of the disease, progressively intensifying and systematically regressing at different age periods, but never go away completely.

General symptoms of chronic inflammatory myopathy include slow but progressive muscle weakness and transverse abdominal disruption. It is marked by frequent constipation.

Eye disorders: pigment retinits, congenital cataracts. Renal failure progresses into a chronic kidney disease between 8–14 years.

The late syndrome is generally noticed between 3 and 6 years of age; it is accompanied by the delayed changes in general medical condition, osteomalacia augmentation and hypokalemic paralysis. Characterized by polyuria and polydipsia, moderate developmental delay, severe gnu valium deformities; low level of phosphate, potassium and calcium in the blood, normal amino acids and glucose concentration. Low plasma bicarbonate concentration is common in the early stages of the disease, later on hyperchloremic acidosis develops. It is characterized by disturbances in the concentration of renal function (hipostenurin, polyuria), sometimes moderate proteinuria, generalized hyperaminociduria, elevated excretion of phosphates, calcium, glucose, citrates in patients. Urine reaction is neutral or alkaline (Lichter-Konecki et al., 2001; Tasic et al., 2008; Watanabe, 2017).

In patients with de Toni-Debre-Fanconi syndrome the following laboratory abnormalities:

- hypophosphatemia;
- hypocalcemia;
- hypokalemia;
- hypernatriemia;
- elevated alkaline serum phosphatase;
- metabolic acidosis (pH: 7.25–7.35; base excess BE (elevated level of alkalinity): –12– –10 mmol/L) secondary to reduced proximal reabsorption of bicarbonate:
  - increased Pyruvic and Lactic Acid Content of Blood;
  - hypophosphatemia;
  - calcitriin;
  - polyuria;
  - decreased serum uric acid with an increased uric acid clearance;
  - glycosuria (above 20–30 g/L);
  - development of generalized hyperaminociduria (less than 2.0–2.5 g/24 h in all the amino acids);
  - failure of amino acid genesis – reduce titratable acidity;
  - increased urine pH (higher than 6.0);
  - tubulin-like proteinuria - the presence of immunoglobulins in the urine of the light chains, lysozyme, β-microglobulin.

Radiological method features has proven to be useful in detecting pronounced osteoporosis with severe disorders in metaphyseal areas
with characteristic bowing of the bones, accompanied by a delayed bone age relative to chronological age of a child.

Additionally, type II is characterized by increased serum 25-hydroxyvitamin D levels in children and decreased – in adults (Levchenko et al., 2006; Magen et al., 2010). In type III, varus angulation of the lower extremities, while renal failure does not occur (Klootwijk et al., 2014). Type IV could be suspected in infants who are large for their gestational age (more than 4 kg), subject to neonatal hypoglycemia, hyperinsulinism and hepatomegaly. There is a risk of development of insulin dependent diabetes mellitus (maternal diabetes of the young (MODY)) accompanied by nephrocalcinosis and renal failure (Hamilton et al., 2014).

**Treatment**

- **Dietary restrictions:**
  - in galactosemia: milk;
  - in fructose intolerance: sugar, honey, apples, pears, watermelons, carrots;
  - in cystinosis: protein foods, high-methionine foods, kitchen salt;
  - in tyrosinemia: high-tyrosine and methionine foods;

- **Recommended foods** (Novikov et al., 2004; Savenkova and Levtchenko, 2004):
  - inpyruvatecarboxylase deficiency: low carb high fat diet (LCHF diet);
  - potassium-, calcium-, phosphorus-rich foods;
  - liquid intake is typically not restricted.

Correction for renal tubular metabolic acidosis:

- 2% or 4% sodium bicarbonate solution (5 ml/kg/day) in 4 divided doses (intravenous, oral, rectal administration) and calcium supplements;
- citrate mixture to reduce the dose of sodium bicarbonate;

- treatment of hyperkalemia (potassium supplements);
- Correction for hyperphosphatemic rickets with normal calcium level and/or hypocalcemia, osteoporosis:
- calcium supplements (calcium carbonate, calcium phosphate, calcium citrate, calcium glycerophosphate). Phosphate buffer (continuously);
- Active metabolites of vitamin D: oxide; calcidiol; calcitriol; or calcium-, phosphorus- and calcitriol- containing binding agents.

Recombinant human growth hormone treated with 0.6-0.7 IU/kg/ week of rhGH administered daily for 3 months.

**Renal tubular acidosis (RTA)**

Several bone deformities in children with tubulopathies are associated with a number of factors: metabolic acidosis should be considered a sign of an underlying disease process. The most vital parameter affecting protein binding of calcium is the pH. Since bone responds to over-acidity, chronic metabolic acidosis of any origin can cause growth retardation. In addition, metabolic acidosis causes alterations in the bone reabsorptive capacity for calcium and therefore increases urinary calcium excretion. The development of metabolic acidosis is caused by a violation of reabsorption of bicarbonates and secretion of hydrogen ions, as well as a violation of the activity of carbonic anhydrase with respect to hydration of CO₂ (this enzyme also stimulates proton secretion not only in renal proximal tubules and collecting ducts, but also in osteoclasts) (Kartarnakisheva et al., 2011).

The disease is inherited by both auto dominant and auto recurrent types; and is clinically characterized by hyperchloremic acidosis and baseline deficiency in the serum.

There are two types of disease: distal renal tubular acidosis (dRTA) type I is characterized by an impairment of the normal urinary acidification process in the distal part of the nephron; Proximal renal tubular acidosis (pRTA) type II is characterized by a defect in the ability to reabsorb bicarbonates in the proximal tubule. Type III is a combination of isolated proximal (type 2) or distal (type 1) tubular pathologies.

Type I dRTA

There are distinguished two types of the disease by the pattern of inheritance: autosomal dominant or autosomal recessive.

**Classical type I dRTA autosomal dominant (OMIM 179800)**

The syndrome is caused by mutations in the SLC4A1 gene (MIM 109270) gene, found in a place on the long arm of chromosome 17 called 17q21.31 (Bergwitz et al., 2006). Clinically it is characterized by osteomalacia, plastic deformity of the long tubular bones and growth retardation. It is caused by the disorder of the tubular acidogenesis, when the kidneys fail to reduce the urine pH associated with the increase in hydrogen ion concentration as a result of the increased reverse diffusion of hydrogen ions through the tight junctions that hold the tubular epithelial cells. The distal canal is unable to create a concentration gradient between the tubular fluid and the blood. This finding suggests that bicarbonate ions have been effectively replaced by chloride ions and the hyperchloremic metabolic acidosis arises (Fry & Karet, 2007; Kraut et al., 2010).

First, in people with this syndrome in their teens or adulthood the following signs and symptoms are observed: poor appetite, polyuria, polydipsia, rapid fatigability and delayed physical development. Next bone deformities commonly associated with rickets (lower-limb valgus deformity, “rachitic rosary”, widening of wrist, frontal and parietal lobe), as well as with the pronounced muscular hypotonia. The first manifestations of the renal tubular acidosis usually appear in children two years of age. People with more severe and prolonged rickets may experience permanent bone deformities (Rodriguez, 2002; Karet, 2002; Civitelli & Zambellas, 2011).

Laboratory studies have revealed metabolic acidosis, low plasma bicarbonate- and increased plasma chloride concentration, hypocalcemia, hypokalemia, increased alkaline phosphatase activity, secondary hyperparathyroidism and decreased intestinal calcium absorption. High-resolution ultrasound has found to be a sensitive and reliable method for the detection of nephrocalcinosis. A significantly decreased renal function (urine specific gravity from 1001 to 1008) is observed, a persistently low urine pH (<5.5), as well as normal bicarbonate levels. Hypercalcitriaemia is associated with excessive urinary calcium excretion (as a compensation for a metabolic acidosis) (Bergwitz & Jäppner, 2010; Escobar et al., 2013) mediated by the renal Ca²⁺ transport proteins (Laing & Unwin, 2006; Nijenhuis et al., 2006) and increased renal sodium reabsorption. This association may implicate increased renal blood flow as a contributory cause of urinary hyperexcretion of insoluble mineral salts, which can lead to recurrent kidney stones or nephrocalcinosis. These factors, together with high urine pH, contribute to abnormal accumulation of calcium and the development of nephrocalcinosis and / or renal stones, which may lead to further deterioration of renal function (Karet, 2002; Loymana et al., 2010).

**Type I autosomal recessive dRTA with deafness or with preserved hearing (OMIM 602772) Syn.: RTADR.**

Defects in the ATP6V0A4 (7q34) or ATP6V1B1 (2p13.3) genes cause autosomal recessive dRTA with deafness and with preserved hearing, respectively. However, several patients with ATP6V0A4 mutations have developed hearing loss, usually in young adulthood.

**Clinical features.** This syndrome occurs in early childhood associated with frequent vomiting and development of dehydration followed by growth retardation and nephrocalcinosis, preceded by chronic renal insufficiency. Often the syndrome is associated with the development of neurosensory deafness. Laboratory findings are the same as in the autosomal dominant form. Parents are usually married (Leung, 2014).

**Type II proximal renal tubular acidosis with ocular abnormalities and mental retardation (OMIM 604278)**

The syndrome is caused by the function mutations in the SLC4A4 gene (MIM 603345) located on chromosome 4 (4q13.3) (Igarashi et al., 2001).

It is associated with the immature nephrons, low carbonic anhydrase II(c) and I(b) activity; as well as low HCO₃⁻ ATPase activity in mitochondria of renal tubular cells.

**Clinical features.** Reduced proximal tubular reabsorption of bicarbonate, resulting in impaired capacity for net acid excretion and persistent hyperchloremic metabolic acidosis. In the first few months of life a history of vomiting, thirst, subëlêre temperature, marked delay in physical growth, rickets-like changes in the skeleton may be present. Developmental delay, myalgias, congenital cataracts, corneal stromal opacities, glaucoma and permanent enamel hypoplasia (Petitfor, 2008).

**Laboratory diagnosis:** increased osmotic fragility of erythrocytes with slightly acid urine (pH less than 6). Hydrogen ions (H⁺) excretion remains within normal limits and corresponds to nutrition. The bicarbonate threshold for bicarbonate reabsorption is decreased, while its excretion is sharply increased.
Treatment: Dietary restriction of oxalate intake (sorrel, spinach, tomato juice, chocolate, etc.), alkaline mineral water, administration of sodium bicarbonate to restore normal acid base status; or diluted citrate solution (Shohla solution) at a dose 5–30 mmol HCO3−/kg/day. Adding more potassium is typically needed. Shohla solution (pharmacy – prepared) containing in 1000 mL not less than 140 g of citric acid and 90 g of sodium citrate (1 g of NaHCO3 = 12 mg of alkaliota, 10 mL of Shohla solution = 10 mg of alkaliota). Vitamin D treatment in patients with osteoporosis and osteomalacia.

Combined proximal and distal renal tubular acidosis (Type III RTA) (Autosomal recessive inheritance associated with osteoporosis) (OMIM 267200)

The syndrome is almost invariably associated with increased bicarbonate excretion.


**Hyrophosphatemic rickets (hypophosphatemia)**

The maintenance of normal phosphate homeostasis constitutes the basic physiologic function of the kidneys (Bastepe & Jüpper, 2008; Natochin, 2008). Serum phosphate concentration exists in three major forms: free ionized (84–85%), protein-bound (10%), and calcium-, magnesium- and sodium compounds (1%) (Escobar et al., 2013). If urine pH is > 7.4, approximately 80% of total phosphate concentration is in the divalent form (HPO42−), while 20% will be in the monovalent form (H2PO4−) (Kartamyshyeva et al., 2011). Usually, about 90% of phosphate in the glomerular filtrate is reabsorbed in the proximal tubule, and 80% reabsorbed proximally (Bastepe & Jüpper 2008; Natochin, 2008; Escobar et al., 2013). The currently known main regulators of phosphate homeostasis include parathyroid hormone (PTH) and vitamin D3 (calcitriol) (Escobar et al., 2013) and leads to a higher plasma phosphorus concentration (Baroncelli et al., 2012).

Calcitriol or biologically active form of vitamin D3 stimulates phosphate reabsorption. Phosphatonin includes fibroblast growth factor 23, frizzled-related protein-4 and phosphoglycoprotein extracellular matrix. Fibroblast growth factor-23 (FGF-23) is a 26-kDa protein activating the specific cell surface receptors FGFRs (Perroud & Portale, 2011).

The ENPP1 (173335), PHEX (300550), DMP1 (600980) and FGF23 genes stimulate the elevation of fibroblast growth factor 23 (FGF-23). The FGF-23 gene is located on chromosome 12p13.3. Fibroblast growth factor 23 (FGF-23) is the gene identified as causative for autosomal dominant hypophosphatemia rickets (Bai et al., 2004; Ben-Dov et al., 2007).

The biological activity and physiological role of FGF-23 have recently been clarified. Several animal models (mice with excess FGF-23 activity as a result of in vivo forced overexpression) exhibit hyperphosphatemia and increased P excretion of 1.25-dihydroxyvitamin D (Sitara, 2004a; Shimada et al., 2004b). FGF-23-deficient mice are characterized by a severe aging-like phenotype associated with ectopic calcifications organ atrophy, and osteomalacia. Mice lacking FGF-23 were characterized by severe vascular- and soft tissue calcification (Kuro-o et al., 1997). Needless to mention that extensive vascular and soft tissue calcification in both FGF-23 and klotho ablated mice are associated with severe hyperphosphatemia, and increased serum level of hydroxvitamin D. The FGF-23 biology was studied on mouse models treated with recombinant FGF-23 or overexpression of FGF-23. FGF-23 suppresses the expression of the types IIa and IIc sodium-phosphate cotransporters on the apical membrane of renal proximal tubular cells, thus inducing phosphaturia (Shimada et al., 2004; Shimada et al., 2009; Shimada et al., 2004a; Shimada et al., 2004b). FGF-23 deficient mice are characterized by a severe aging-like phenotype associated with ectopic calcifications organ atrophy, and osteomalacia. Mice lacking FGF-23 were characterized by severe vascular- and soft tissue calcification (Kuro-o et al., 1997). Needless to mention that extensive vascular and soft tissue calcification in both FGF-23 and klotho ablated mice are associated with severe hyperphosphatemia, and increased serum level of hydroxvitamin D. The FGF-23 biology was studied on mouse models treated with recombinant FGF-23 or overexpression of FGF-23. FGF-23 suppresses the expression of the types IIa and IIc sodium-phosphate cotransporters on the apical membrane of renal proximal tubular cells, thus inducing phosphaturia (Shimada et al., 2005). The phosphatidic action of FGF-23 is not expressed in the absence of sodium-hydrogen exchanger regulatory factor-1 (NHERF-1) and increases in the presence of parathyroid hormone (PTH). In addition, FGF-23 suppresses the formation of 1,25(OH)2D, suppressing 1-alpha-hydroxylase (CYP27B1), which converts 25-hydroxyvitamin D [25(OH)D] to 1,25(OH)2D and stimulates the formation 24-hydroxylase (CYP24), which converts 1,25(OH)2D into inactive metabolites in the proximal tubule of the kidneys. In addition, FGF-23 impairs the production of renal 1,25-dihydroxyvitamin D [1,25(OH)2D] by inhibiting the expression of CYP27B1, the enzyme that converts 25-(OH)D to inactive metabolites in the proximal tubule. FGF23 also reduces the expression of interstitial sodium-phosphate cotransporter NPT2b (Saito et al., 2003), reducing the intestinal phosphate absorption.

FGF-23 acts directly on the parathyroid gland to inhibit PTH synthesis and secretion. It has been shown that FGF-23 activates the mitogen-activated protein kinase pathway leading to a decrease in parathyroid hormone (PTH) secretion both in vivo rats and in vitro shingles (Ben-Dov et al., 2007). FGF-23 has also been shown to increase expression of parathyroid 1-alpha-hydroxylase (Krajisnik et al., 2007), which converts 25-hydroxyvitamin D [25(OH)D] to 1,25(OH)2D.

FGF-23 secretion is regulated by local bone-derived factors, such as phosphate-regulating gene with homologies to endopeptidases and den- tin matrix protein-1 (Lorenz-Dcapecreux et al., 2006a; Lorenz-Dcapecreux et al. 2006b). 1,25(OH)2D affects FGF-23 secretion both in vivo and in vitro through the activation of FGF-23 mediated vitamin D (Liu et al., 2006).

1. Factors decreasing phosphate reabsorption:
   - parathyroid hormone;
   - atrial natriuretic peptide;
   - glucocorticoids;
   - dopamine.

2. Factors increasing phosphate reabsorption:
   - parathyroidecocy;
   - 11α25(OH)2D3;
   - growth hormone;
   - insulin-like growth factor;
   - food regulation;
   - acute (minutes, hours);
   - chronic (hours, days);
   - several system factors.

X-linked recessive hypophosphatemia rickets (Dent’s disease) (OMIM 300554)

The disease is caused mainly by mutations in the CLCN5 gene located on chromosome Xp11.22.

X-linked recessive hypophosphatemia rickets is a form of X-linked hypercalciuric nephro lithiasis, which comprises a group of disorders characterized by proximal renal tubular reabsorptive failure, hypercalciuria, nephrocalcinosis, and renal insufficiency.

Clinical features: rickets or osteomalacia, hypercalciuria, hypophosphatemia and proteinuria in children. Progressive calcification and renal failure in adult patients (Garnбаро et al., 2004). Clinically, it may show bone pain, fatigue, muscle weakness, and repeated bone fractures. Symptoms are related to bone pain, fatigue, muscle weakness and recurrent bone fractures.

Autosomal dominant hypophosphatemia rickets (OMIM 193100)

Autosomal dominant hypophosphatemia rickets (ADHR) results from activating mutations in a fibroblast growth factor 23 (FGF-23) gene in chromosome 12p13 encoding a phosphate-regulating hormone (Sun et al., 2012; Wöhrle et al., 2013).

Small amounts of the gene originate in the brain, thymus, small intestine, heart, liver, lymph nodes, thyroid-shaped and pterygoid glands, bone marrow and in large quantities in tumors with oncogenic osteomalacia. No expression in the bones. Elevated levels of FGF-23 are associated with inhibition of reabsorption of phosphates in the renal tubule and hypophosphatemia. FGF-23 can physiologically function as a locally active factor secreted in excess amounts in conditions of pathology, and may cause renal phosphate loss.

Less than 100 cases have been described.

Clinical manifestations depend on the age of onset and on the severity of hypophosphatemia.

Clinical features: ADHR shows incomplete penetrance and variable age at onset (childhood to adult). Phosphate excretion can be evaluated by measuring the maximum tubular reabsorption per glomerular filtration rate.

Laboratory diagnosis: It is characterized by severe hypophosphatemia arising from a defect in the renal reabsorption of filtered inorganic phosphate (Pi), elevated serum alkaline phosphatase activity and fibroblast growth factor 23 (FGF-23), inappropriately low-normal serum concentration of 1,25-dihydroxyvitamin D3 [1,25(OH)2D3] levels for the degree of prevailing hypophosphatemia (Econs et al., 1997).

Treatment: aimed at improving growth, enhancing mineralization of bones, and preventing skeletal deformities caused by rickets. It consists of daily oral administration of phosphate and calcitriol and is associated with frequent monitoring of calcium, alkaline phosphatase, and parathyroid hormone, and phosphate serum concentrations, as well as urinary calcium and creatinine.

Autosomal recessive hypophosphatemia rickets type 1 (ARHR1) (OMIM 241530)

ARHR1 is caused by homozygous loss-of-function mutations in the DMP1 (Dentin matrix protein 1) gene in chromosome 4q22.

Clinical features: Lower-extremity deformities. No response to vitamin D therapy (vitamin D resistant rickets), high bone density. Back pain, restricted joint motion. Premature fusion of the skull bones. Deafness (aplasia of the vestibulocochlear nerve that results in ipsilateral congenital sensorineural hearing loss). Dental defects and early caries. It is accompanied by muscle weakness and pathologic fractures.

Radiographic evaluation: early osteosclerosis and skull thickening, trabecular bone density in ribs (Feng et al., 2006; Lorenz-Depiereux et al., 2006a).

Laboratory diagnosis: there are no symptoms of hypophosphatemia. Autosomal recessive hypophosphatemia rickets type 2 (ARHR2) (OMIM 613312)

ARHR2 is caused by homozygous loss-of-function mutation in the ENPP1 gene in chromosome 6 (q6). Mutations in ENPP1 gene are also responsible for generalized arterial calcification of infancy.

Clinical features: hypophosphatemia rickets, sometimes generalized arteriolar calcification of infancy (Lorenz-Depiereux et al., 2010).

Laboratory diagnosis: hypophosphatemia.

X-linked, dominant, hypophosphatemia rickets (XLHR) (OMIM 307380)

Inactivating mutations in PHEX gene with homologies to endopeptidase on the X chromosome (Xp22) have been identified as a cause of XLHR. This endopeptidase is mainly expressed in bones and teeth, regulating FGF-23 synthesis. The disease occurs as an X-linked dominant disorder with complete penetrance often complicated by variable expressivity.

PHEX revealed possible alternative regulatory mechanisms for phosphate homeostasis, bone mineralization, and vitamin D metabolism. It controls sodium-dependent phosphate transport proteins in intestinal and renal proximal tubular epithelial cells. The genetic disorder is associated with inability of the renal proximal tubule to reabsorb phosphate, which affects intestinal phosphate absorption. PHEX is primarily expressed in osteoblasts, odontoblasts, lung, ovary, parathyroid gland, brain and muscle. We found no correlation between the location or type of mutation and the disease severity.

In XLHR osteoblast is likely to produce some inhibitor. Moreover, it was reported that cross-transplantation of kidneys in hyp-children results in transfer of the mutant phenotype. It can be associated with the primary defect in osteoblasts, as the correction of hyperphosphatemia and calcitriol in patients were observed low mineralization zones around osteocytic lacunae. XLH is the most frequent form of hypophosphatemia rickets, with a prevalence of 1/20,000. The disease affects both sexes. Patients with early onset disease have phosphate wasting, rickets, and lower extremity deformities in childhood.

Characteristics heritable dental developmental anomalies: enamel hypoplasia, dentinogenesis imperfecta, enlarged dental tubules, leading to tooth abscesses. Characteristic cranial base abnormalities: thickening of outer cortical table of frontal bone and slightly sunken median line between the eyes at the forehead. Osteocartilaginous of the lower extremities is developed in adults, osteophytes are formed and in some cases hearing loss may occur. Muscular weakness and hypotension are not observed. Other clinical manifestations, such as enthesisopathy (calcification of ligaments and their attachment to bone), which is accompanied by joint pain and joint mobility disorders (Baroncelli et al., 2012).

Laboratory diagnosis: hypophosphatemia with low renal phosphate reabsorption, normal serum calcium values, normal or low vitamin D serum level (1,25(OH)2D3 or calcitriol), normal serum parathyroid hormone levels and increased serum alkaline phosphatase activity.

Therapy aimed at normalization of PTH levels with calcitriol supplementation and calcitriol. Correction of vitamin D deficiency state is not required. Calcitriol supplementation may prevent secondary hyperparathyroidism resulting from increased phosphate intake (Gaucher et al., 2009).

Conclusions

Although the diagnosis of tubulopathy remains complex and requires expensive laboratory equipment and specialist expertise; it can be diagnosed in children showing the following symptoms: impaired growth, vitamin D resistant rickets (lower limb deformities between 2 and 3 years of age). In the evaluation of such patients urine analysis is commonly used (levels of calcium, phosphorus, pH, bicarbonate, sodium, potassium, glucose, creatinine, protein, amino acids), blood count (levels of creatinine, uric acid, alkaline phosphatase, glucose, pH and sodium, bicarbonate, potassium, chloride, calcium, phosphorus ions) and ultrasound of the kidneys to detect nephrocalcinosis. Determination of serum parathyroid hormone concentration, vitamin D metabolites, aldosterone and plasma renin activity, cysteine lymphocyte concentration and urothelial examination may also be used as additional diagnostic methods. Despite the fact that most tubulopathies can be diagnosed clinically, molecular genetic studies are needed to clarify the type of inheritance and prognosis.

References


and identifies a role for osteocytes in mineral metabolism. Nature Genetics, 38(11), 1310–1315.


